A Colorimetric and Ratiometric Fluorescent Probe for Palladium

Jie Jiang, Huie Jiang, Wei Liu, Xiaoliang Tang, Xi Zhou, Weisheng Liu,* and Ruiting Liu

Key Laboratory of Nonferrous Metals Chemistry and Resources Utilization of Gansu Province and State Key Laboratory of Applied Organic Chemistry, College of Chemistry and Chemical Engineering, Lanzhou University, Lanzhou 730000, P. R. China

liuws@lzu.edu.cn

Received July 25, 2011



A colorimetric and ratiometric fluorescent probe for the palladium species has been developed based on the Pd⁰-catalyzed cleavage of an allyoxycarbonyl group of amines under mild conditions. The probe displays a highly sensitive and selective response with significant changes in both color (from colorless to jade-green) and fluorescence (from blue to green), through the ICT process.

With the wide use of the palladium species in pharmacy and various materials,^{1,2} the resulting high level of residual palladium in the final product has raised great concern.³ Palladium can have adverse effects on our health and the environment, because it can bind to thiol-containing amino acids, proteins, DNA, and other biomolecules and disturb a variety of cellular processes.¹ Thus, the final threshold for palladium in the end products is strictly limited, with set governmental restrictions (no more than 5-10 ppm) and a proposed maximum dietary intake of palladium (as a crude estimate) of less than $1.5-15 \mu g$ per person per day.⁴ This issue raises the urgent need to develop effective methods for palladium species sensing.

Conventional analytical methods for palladium species include atomic absorption spectrometry (AAS), inductively coupled plasma mass spectrometry (ICP-MS), solid phase microextraction-high performance liquid chromatography, X-ray fluorescence, etc.⁵ However, these methods

LETTERS 2011 Vol. 13, No. 18 4922–4925

ORGANIC

⁽¹⁾ International Programme on Chemical Safety. Palladium; Environmental Health Criteria Series 226; World Health Organization, Geneva, 2002.

^{(2) (}a) Lyons, T. W.; Sanford, M. S. Chem. Rev. 2010, 110, 1147–1169. (b) Cho, D. G.; Sessler, J. L. Chem. Soc. Rev. 2009, 38, 1647–1662.
(c) Eryazici, I.; Moorefield, C. N.; Newkome, G. R. Chem. Rev. 2008, 108, 1834–1895. (d) Buchwald, S. L.; Mauger, C.; Mignani, G.; Scholz, U. Adv. Synth. Catal. 2006, 348, 23–39. (e) Zeni, G.; Larock, R. C. Chem. Rev. 2004, 104, 2285–2310. (f) Tietze, L. F; Ila, H.; Bell, H. P. Chem. Rev. 2004, 104, 3453–3514.

^{(3) (}a) Cho, D. G.; Sessler, J. L. *Chem. Soc. Rev.* 2009, *38*, 1647–1662.
(b) Kielhorna, J.; Melber, C.; Keller, D.; Mangelsdorf, I. *Int. J. Hyg. Environ. Health* 2002, *205*, 417–432.

⁽⁴⁾ Garrett, C. E.; Prasad, K. Adv. Synth. Catal. 2004, 346, 889–900.
(5) (a) Van Meel, K.; Smekens, A.; Behets, M.; Kazandjian, P.; VanGrieken, R. Anal. Chem. 2007, 79, 6383–6389. (b) Locatelli, C.; Melucci, D.; Torsi, G. Anal. Bioanal. Chem. 2005, 382, 1567–1573. (c) Dimitrova, B.; Benkhedda, K.; Ivanova, E.; Adams, F. J. Anal. At. Spectrom. 2004, 19, 1394–1396.

^{(6) (}a) Kim, J. S.; Quang, D. T. *Chem. Rev.* **2007**, *107*, 3780–3799. (b) Angell, S. E.; Rogers, C. W.; Zhang, Y.; Wolf, M. O.; Jones, W. E., Jr. *Coord. Chem. Rev.* **2006**, *250*, 1829–1841. (c) Rogers, C. W.; Wolf, M. O. *Coord. Chem. Rev.* **2002**, *233–234*, 341–350.

⁽⁷⁾ Fluorescent chemosensors for the palladium species: (a) Goswami, S.; Sen, D.; Das, N. K.; Fun, H.; Quah, C. K. *Chem. Commun.* **2011**, *47*, 9101– 9103. (b) Kim, H.; Moon, K.-S.; Shim, S.; Tae, J. *Chem.—Asian J.* **2011**, *6*, 1987–1991. (c) Li, H.; Fan, J.; Du, J.; Guo, K.; Sun, S.; Liu, X.; Peng, X. *Chem. Commun.* **2010**, *46*, 1079–1081. (d) Li, H.; Fan, J.; Song, F.; Zhu, H.; Du, J.; Sun, S.; Peng, X. *Chem.—Eur. J.* **2010**, *16*, 12349–12356. (e) Schwarze, T.; Dosche, C.; Flehr, R.; Klamroth, T.; Löhmannsröben, H.-G.; Saalfrank, P.; Cleve, E.; Buschmanne, H.-J.; Holdt, H.-J. *Chem. Commun.* **2010**, *46*, 2034– 2036.

often need serious sample-preparation steps or expensive equipment. Thus, current research has been focused on fluorescent methods, because of their low cost, simplicity, and sensitivity.⁶ Some fluorescent chemosensors⁷ and chemodosimeters⁸ for the palladium species have been reported in the literature recently. To the best of our knowledge, most of these sensors respond to palladium with changes only in fluorescence intensity (based on the ON-OFF⁹ or OFF-ON mechanism¹⁰). However, a major limitation of the intensity-based sensor is that quantitative detection can be significantly influenced by the excitation power and detector sensitivity.¹¹ In contrast, ratiometric fluorescent measurement which uses the ratio of two fluorescent bands instead of the absolute intensity of one band, makes measuring the analyte more accurately and sensitively possible with minimization of the background signal.¹² Herein, we developed a ratiometric fluor-





escent probe for the palladium species with a perceived color change.

(10) (a) Santra, M.; Ko, S. K.; Shin, I.; Ahnz, K. H. Chem. Commun.
2010, 46, 3964–3966. (b) Garner, A. L.; Koide, K. J. Am. Chem. Soc.
2008, 130, 16472–16473. (c) Song, F.; Garner, A. L.; Koide, K. J. Am. Chem. Soc.
2007, 129, 12354–12355. (d) Schwarze, T.; Müller, H.; Dosche, C.; Klamroth, T.; Mickler, W.; Kelling, A.; Lömannsröen, H.-G.; Saalfrank, P.; Holdt, H.-J. Angew. Chem., Int. Ed. 2007, 46, 1671–1674.

(13) (a) Xu, Z.; Yoon, J.; Spring, D. R. *Chem. Commun.* **2010**, *46*, 2563–2565. (b) Zhu, B.; Zhang, X.; Li, Y.; Wang, P.; Zhang, H.; Zhuang, X. *Chem. Commun.* **2010**, *46*, 5710–5712. (c) He, H.; Mortellaro, M. A.; Leiner, M. J. P.; Fraatz, R. J.; Tusa, J. K. *J. Am. Chem. Soc.* **2003**, *125*, 1468–1469.

In our scaffold, 4-aminonaphthalimide was chosen as the fluorophore because of its outstanding ICT structure^{12c,13} and desirable photophysical properties, such as a large Stokes' shift, long emission wavelength, and insensitivity to pH (Scheme 1). Compound 1, *N*-butyl-4-NHAlloc-1,8-naphthalimide, was prepared smoothly from *N*-butyl-4-amido-1,8-naphthalimide (2) and allyl chloroformat in a satisfactory yield. It can be easily cleaved with a quantitative conversion to the free amino compound 2 after being treated with Pd(PPh₃)₄ under mild reaction conditions.¹⁴



Figure 1. Fluorescence spectral changes of probe $1 (20 \,\mu\text{M})$ upon treatment with Pd(PPh₃)₄ (10 μ M), in acetonitrile–water solutions (CH₃CN:H₂O = 4:1, NaBH₄–PPh₃ (10 mM) and morpholine (10 mM)) at room temperature. Ex = 403 nm. Slit: 5.0 nm/5.0 nm. Inset: Photos showing (a) color and (b) fluorescence color changes upon addition of Pd(PPh₃)₄.

The spectroscopic properties of the compound 1 were rated in acetonitrile–water solutions ($CH_3CN:H_2O = 4:1$, $NaBH_4$ –PPh₃ (10 mM) and morpholine (10 mM)) at a micromolar concentration. As shown in Figure 1, free compound 1 (20 μ M) showed a relatively short emission wavelength with a maximum at 462 nm (blue), owing to the electron-withdrawing effect of the amide group. After being treated with $Pd(PPh_3)_4$ (10 μ M), the maximum emission peak underwent a red shift to 524 nm (green), showing a ratiometric response. This red shift in the emission should be attributed to the stronger ICT efficiency of the released amino compound 2 (Figure S1). A red shift from 370 to 430 nm of 1 was also detected in the maximum absorption spectrum. Additionally, marked color changes in the solutions, which ranged from colorless to jade-green, could be distinguished by the naked eyes. Clearly, the results indicated that Pd⁰ could be detected through both ratiometric fluorescence and colorimetric methods by probe 1.

To explore the reactivity of probe 1 toward other palladium metal sources, different initial oxidation states of palladium such as PdCl₂, Pd(OAc)₂, Pd(CH₃CN)₂Cl₂, and Pd(PPh₃)₂Cl₂ were examined (Figure S5). To our delight, all the samples gave similar responses to probe 1 as Pd(PPh₃)₄, indicating the response depends on palladium itself, regardless of its ligands or complex anions.

⁽⁸⁾ Fluorescent chemodosimeters for the palladium species: (a) Song, F. L.; Carder, E. J.; Kohler, C. C.; Koide, K. *Chem.—Eur. J.* 2010, *16*, 13500–13508. (b) Jun, M. E.; Ahn, K. H. *Org. Lett.* 2010, *12*, 2790–2793. (c) Schwarze, T.; Mickler, W.; Dosche, C.; Flehr, R.; Klamroth, T.; Lömannsröen, H.-G.; Saalfrank, P.; Holdt, H.-J. *Chem.—Eur. J.* 2010, *16*, 1819–1825. (d) Garner, A. L.; Song, F.; Koide, K. *J. Am. Chem. Soc.* 2009, *131*, 5163–5171. (e) Garner, A. L.; Koide, K. *Chem. Commun.* 2009. 86–88.

^{(9) (}a) Liu, B.; Bao, Y.; Du, F.; Wang, H.; Tian, J.; Bai, R. Chem. Commun. 2011, 47, 1731–1733. (b) Duan, L.; Xu, Y.; Qian, X. Chem. Commun. 2008, 6339–6341. (c) Matthews, J. R.; Goldoni, F.; Kooijman, H.; Spek, A. L.; Schenning, A. P. H. J.; Meijer, E. W. Macromol. Rapid Commun. 2007, 28, 1809–1815. (d) Tamayo, A.; Escriche, L.; Casabo, J.; Covelo, B.; Lodeiro, C. Eur. J. Inorg. Chem. 2006, 2997–3004. (e) Unterreitmaier, E.; Schuster, M. Anal. Chim. Acta 1999, 378–402, 339–344. (f) Kubo, K.; Miyazaki, Y.; Akutso, K.; Sakurai, T. Heterocycles 1999, 51, 965–968.

^{(11) (}a) Srikun, D. E.; Miller, W.; Domaille, D. W.; Chang, C. J. *J. Am. Chem. Soc.* **2008**, *130*, 4596–4597. (b) Komatsu, K.; Urano, Y.; Kojima, H.; Nagano, T. *J. Am. Chem. Soc.* **2007**, *129*, 13447–13454.

^{(12) (}a) Zhu, B.; Gao, C.; Zhao, Y.; Liu, C.; Li, Y.; Wei, Q.; Ma, Z.; Du, B.; Zhang, X. Chem. Commun. 2011, 47, 8656–8658. (b) Domaille, D.; Zeng, W. L.; Chang, C. J. J. Am. Chem. Soc. 2010, 132, 1194–1195. (c) Xu, Z.; Baek, K. H.; Kim, H.; Cui, J.; Qian, X.; Spring, D.; Shin, I.; Yoon, J. J. Am. Chem. Soc. 2010, 132, 601–610. (d) Ajayaghosh, A.; Carol, P.; Sreejith, S. J. Am. Chem. Soc. 2005, 127, 14962–14963.

⁽¹⁴⁾ For detailed experimental procedures and the characterization of the new compounds, please see the Supporting Information.

Since $PdCl_2$ is the most toxic species, among the palladium compounds,^{10a} we proceeded to study the detection of $PdCl_2$ in the following experiment.



Figure 2. Fluorescence spectra of probe **1** upon titration of Pd²⁺ $(0-2 \mu M)$ in acetonitrile – water solutions (CH₃CN:H₂O = 4:1, NaBH₄–PPh₃ (10 mM) and morpholine (10 mM)), taken after 5 min at room temperature. Ex = 403 nm. Slit: 5.0 nm/5.0 nm. [probe **1**] = $20 \mu M$. Inset: Ratiometric calibration curve $I_{524 nm}/I_{498 nm}$ as a function of Pd²⁺.

Upon titration of Pd²⁺ (Figure 2), a gradual decrease in fluorescence intensity at 462 nm and the simultaneous appearance of a new red-shifted emission band at 524 nm were observed with an isoemission point at 498 nm, indicating a clear ratiometric fluorescence change. The fluorescence intensity ratios of probe 1 at 524 and 498 nm ($I_{524 \text{ nm}}/I_{498 \text{ nm}}$) increased linearly with the amount of Pd²⁺ in the range of 0–1 μ M (Figure 2, inset). The detection limit of probe 1 for PdCl₂ was determined as 6.1 nM (Pd content = 0.65 μ g/L) (see the Supporting Information), much lower than the palladium content in persons found from samples of morning saliva (10.6 \pm 7.4 μ g/L).¹ That is, probe 1 can be a sensitive ratiometric fluorescent sensor for the quantitative detection of Pd²⁺.

To demonstrate the practical application of the probe, we carried out an experiment to monitor residul Pd in a reactor using probe 1. A THF solution of PdCl₂ was stirred in four flasks for 1 h at room temperature. After the



Figure 3. Color changes in probe 1 (20 μ M) treated with the residual PdCl₂ on the surface of glassware. Left: Visible color on excitation at 365 nm using a hand-held UV lamp; Right: Visual fluorescence. From 1 to 4: Probe 1 solutions not exposed to PdCl₂ reagents (1) and exposed to PdCl₂ on the surface of glassware with different wash procedures (brushing with detergent, washing with water and acetone (2); brushing with detergent and washing with water (3); and brushing with detergent only (4)).

solution was poured out, the four flasks were treated with different washing procedures, respectively (no wash; brushing with detergent only; brushing with detergent and washing with water; and brushing with detergent, washing with water and acetone). Next, probe **1** solution (CH₃CN:H₂O = 4:1, NaBH₄-PPh₃ (10 mM) and morpholine (10 mM)) was added into the four flasks. After 5 min, obvious fluorescent and color changes were detected (Figure 3). Thus, probe **1** could be used for the quality control of reactors through convenient visual sensing.

Furthermore, a proof-of-concept experiment for Pd²⁺ detection in the environment was also performed with pool and tap water. The water samples were collected and filtered, prepared as acetonitrile–water solutions (CH₃CN:H₂O = 4:1, NaBH₄–PPh₃ (10 mM) and morpholine (10 mM)), and then pretreated with different amounts of PdCl₂ (0–1 μ M). After the addition of probe 1 (20 μ M), PdCl₂ could be readily detected with a large fluorescence signal which was almost linearly dependent on the concentration of palladium (Figure 4). The results demonstrate that our detection system could function well in quantitative Pd²⁺ analysis in environmental samples.



Figure 4. Proof-of-concept experiment with probe 1 (20 μ M) for Pd²⁺ detection in acetonitrile–tap water ($\mathbf{\nabla}$) and acetonitrile–pool water ($\mathbf{\Phi}$) solutions at μ M levels. Ex = 403 nm. Slit: 5.0 nm/ 5.0 nm.

The selectivity experiment was conducted using different metal ions (Figure 5). Under modified conditions¹⁵ in acetonitrile–water solutions (CH₃CN:H₂O = 4:1, PPh₃ (10 mM) and morpholine (10 mM)), the ratiometric response was detected only in the case of Pd²⁺; other metal species such as Au³⁺, Ag⁺, Ru³⁺, Cr³⁺, Mn²⁺, Fe³⁺, Co²⁺, Ni²⁺, Cu²⁺, Zn²⁺, Cd²⁺, and Hg²⁺ had no or little effect on the emission of probe 1. Remarkably, as one that coexists with Pd²⁺ in ores and also competes with it in the sensing studies,^{10b} even Pt²⁺ did not show any influence on our sensor. Moreover, compared with various metal species

⁽¹⁵⁾ Due to the similar π -electrophilicity between cationic Pd and Pt species, Pt²⁺ could also respond to probe 1 in acetonitrile–water solutions (CH₃CN:H₂O = 4:1, NaBH₄–PPh₃ (10 mM) and morpholine (10 mM) (Figure S9). In order to avoid the influence of Pt²⁺, we optimized the conditions, in which NaBH₄ was deleted and the reaction time was prolonged (CH₃CN:H₂O = 4: 1, PPh₃ (10 mM) and morpholine (10 mM)). In this case, the sensor showed specific selectivity towards Pd²⁺.



Figure 5. (a) Fluorescence spectra of probe 1 in the absence and presence of different metal ions Cr^{3+} , Mn^{2+} , Fe^{3+} , Co^{2+} , Ni^{2+} , Cu^{2+} , Zn^{2+} , Cd^{2+} , Hg^{2+} , Ag^+ , Pt^{2+} , Au^{3+} , Ru^{3+} , and Pd^{2+} (as their ClO_4^- or Cl^- salts) in acetonitrile–water solutions (CH₃CN:H₂O = 4:1, PPh₃ (10 mM) and morpholine (10 mM)), measured after 30 min; (b) Metal ion selectivity of probe 1 ($R = I_{524 \text{ nm}}/I_{498 \text{ nm}}$). Inset: Fluorescence changes excited by UV lamp (365 nm) and color changes in probe 1 upon addition of various metal cations. Ex = 403 nm. Slit: 5.0 nm/5.0 nm. [probe 1] = 20 \,\mu\text{M}, [M^{n+}] = 10 μ M.

tested, the only color change in probe **1** with Pd²⁺ made it convenient to detect the palladium species through colorimetric method.

Different anion-induced influences on the sensing behavior of the probe $1-Pd^{2+}$ system were also investigated. As shown in Figure S7, the sensing for Pd^{2+} hardly experienced interference from commonly coexistent anions, such as F⁻, Cl⁻, Br⁻, I⁻, CNS⁻, HSO₄⁻, H₂PO₄⁻, NO₃⁻, ClO₄⁻, and AcO⁻. Furthermore, competition experiments of metal ions showed that the addition of various metal ions promoted a negligible effect on Pd²⁺ sensing (Figure S8).

Scheme 2. Mechanism for Selective Recognition of Palladium



The excellent selectivity should be attributed to the highly specific Pd⁰-triggered cleavage process. A feasible fluorescence sensing mechanism is shown in Scheme 2.¹⁶ Pd⁰ reacts with the allyl carbamate group of compound **1** to yield π -allylpalladium(II) complex **3**, further transferring the allyl unit to morpholine as the nucleophile, and then producing carbamate **4**. Finally, decarboxylation of the compound delivers the compound **2** with the green fluorescene. In this case, the sensor could operate through the cleavage reaction triggered by Pd⁰ with fluorescene changes at two different wavelengths. Furthermore, Pd²⁺ can also be detected by the same principle, when a reducing agent, such as NaBH₄–PPh₃, is added.

In conclusion, we have rationally developed a sensitive and selective fluorescent probe for the palladium species based on the Pd^0 -triggered cleavage reaction under mild conditions. Probe 1 displayed specific and ratiometric fluorescent responses toward palladium, with a marked color change from colorless to jade-green. Moreover, with the ratiometric fluorescent signal and a low detection limit, the probe may be favorable for the quantitative detection of palladium in environmental and chemical settings.

Acknowledgment. This study was supported by the NSFC (Grants 20931003, J0730425) and the Fundamental Research Funds for the Central Universities (lzujbky-2009-k06, lzujbky-2009-114).

Supporting Information Available. Experimental details, characterization for the compounds, and additional spectroscopic data are reported. This material is available free of charge via the Internet at http://pubs.acs.org.

^{(16) (}a) Trost, B. M. Acc. Chem. Res. **1980**, *13*, 385–393. (b) Tsuji, J. Organic Synthesis with Palladium Compounds; Springer-Verlag; Berlin, 1980; pp 125–132. (c) Dangles, O.; Guibé, F.; Balavoine, G.; Lavielle, S.; Marquet, A. J. Org. Chem. **1987**, *52*, 4984–4993.